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The effect of sewage sludge containing microplastics on growth and fruit development of tomato plants

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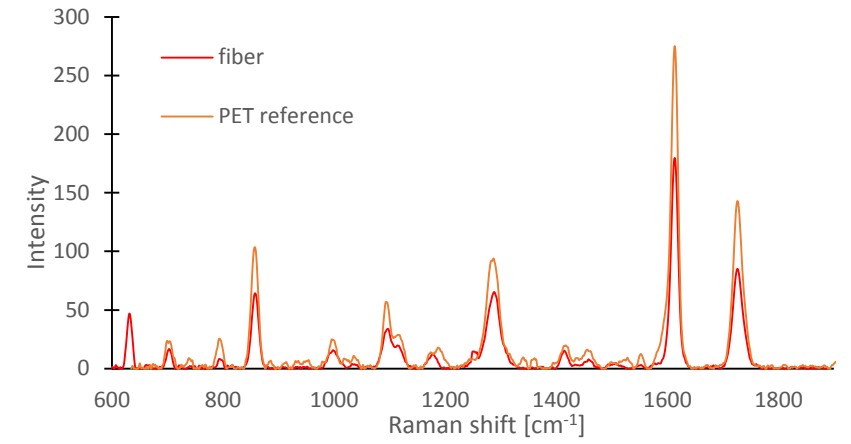
Extraction of the microplastics from sewage sludge



Cultivation of tomato plants with sewage sludge



Characterization and quantification



Measurement of morphological parameters, biomass and fruit production



25 **ABSTRACT**

26 Microplastics (MPs) are becoming an environmental growing concern, being the
27 sewage sludge applied to agriculture fields one of the most important inputs to the
28 environment. To date, there is no standardized protocol for their extraction and changes
29 in vegetative growth and fruit maturation on cultivated plants induced by sludge
30 containing MPs have not been studied yet. Sewage sludge from three different
31 wastewater treatment plants located in Murcia, Spain, were studied. First, the
32 microplastic concentration was estimated and, then, the effects of the sewage sludge in
33 the development of tomato plants and fruit production was analyzed. The measured
34 parameters in tomato plants were both, biomass and length, for shoot and root part, as
35 well as, stem diameter and tomato production. The present work has developed and
36 validated a protocol for the extraction and quantification of MPs comprising several
37 shapes, materials and sizes from samples of sewage sludges, which offers a good
38 compromise for the extraction of different types of microplastic. The protocol used for
39 MPs extraction had a recovery efficiency of 80 ± 3 % (mean \pm SE) and used
40 bicarbonate, to maximize MPs extraction. The mean abundance of MPs in the studied
41 sewage sludge samples was $30,940 \pm 8,589$ particles kg^{-1} dry weight. Soils with sludge
42 containing MPs fostered the growth of tomato plants, while delaying and diminished
43 fruit production. However, other factors or their interactions with MPs could have
44 influenced the outcomes. Further studies are necessary to corroborate these findings and
45 explain the mechanisms of possible effects of MPs on plants.

46 **Capsule:** In this work, the effects of sewage sludges containing microplastics on growth
47 and fruit development in tomato plants have been evaluated.

48

49 **KEYWORDS:** Tomato plants, sewage sludge, microplastics, biomass, fruit crop

50 **1. Introduction**

51 Microplastics (MPs) are an environmental growing concern, with an annual input to
52 the environment of 11 million tons (Boucher and Friot, 2017). Agriculture fields
53 receiving sludge from sewage treatment plants as fertilizers constitute one of the largest
54 inputs of MPs to the environment (63000-430000 t of MPs per year; (Nizzetto et al.,
55 2016). Only in Europe, wastewater treatment plants (WWTPs) produce 13 million tons
56 of dry weight sludge per year and approximately 99% of MPs can be retained in the
57 sludge (Magnusson and Norén, 2014). 80% of this sludge is used in agriculture (Mahon
58 et al., 2017). Despite the importance of quantifying the concentrations of MPs in sludge
59 (de Souza et al., 2018; Corradini et al., 2019), to date, there is no standardized protocol
60 for their extraction and quantification (Rocha-Santos and Duarte, 2015; Rochman et al.,
61 2017).

62 Different methodologies have been proposed for the extraction of MPs from sludge
63 samples, generally using oxidizing agents and acids to remove part of the organic matter
64 and a density separation of the MPs by means of saline solutions (Lusher et al., 2017;
65 Sujathan et al., 2017; Hurley et al., 2018; Lares et al., 2018; Li et al., 2018). However,
66 one of the drawbacks of using high concentrations of these agents to remove organic
67 matter is the possible degradation of some polymers, (Avio et al., 2015), difficulting
68 their identification by using Raman and FT-IR spectroscopy (Lares et al., 2018).

69 Additionally, estimates of MPs concentrations must be accompanied by a recovery
70 test of the most abundant types of MPs comprising different sizes. Previous studies
71 quantifying MPs in sludge or in soil samples have generally performed partial recovery
72 tests only using one (Mahon et al., 2017; Sujathan et al., 2017; Li et al., 2018) or two
73 shapes of MPs (Hurley et al., 2018; Corradini et al., 2019) and in some cases a recovery
74 test was not applied (Lusher et al., 2017; van den Berg et al., 2020). Nevertheless,

75 recovery tests are key in assessing the effectiveness of the extraction methods,
76 necessary to design standardized procedures that can precisely estimate MPs
77 concentrations in sewage sludge.

78 MPs are expected to alter the biophysical properties of the soil, in terms of water
79 retention capacity, bulk density, microbial activity and formation of aggregates, which
80 could lead to an alteration in the water cycle in the soil (de Souza Machado et al., 2018;
81 Wan et al., 2019). However, the effect that MPs accumulation in the soil will generate
82 on the vegetation has been scarcely studied. MPs present in sludge from WWTPs used
83 in agriculture can remain in soil for a long-time affecting the vegetative growth (van den
84 Berg et al., 2020). In this sense, it has been reported that MPs can induce changes in
85 both aerial and below-ground parts during vegetative growth in the wheat plant (Qi et
86 al., 2018) while reducing the germination rate and root growth of *Lepidium sativum* as
87 the concentration of MPs to which plants were exposed increased (Bosker et al., 2019).
88 Although sewage sludge has been employed to improve soil fertility in agriculture
89 (Llorens et al., 2012), to the best of our knowledge, changes induced by sludge
90 containing MPs in vegetative growth and fruit maturation of cultivated plants have not
91 been studied.

92 The aim of this work is to assess the efficiency of a protocol for MPs extraction and
93 quantification from sewage sludge samples and evaluate the effect of sewage sludge
94 containing MPs on tomato plants once applied in soil. To assess the efficiency of the
95 extraction and quantification protocol, a recovery test was carried out using different
96 types of particles, materials and sizes. At last, the effects of sewage sludge containing
97 MPs on tomato plants were evaluated by means of response variables related to growth
98 and fruit production.

99

100

101 **2. Materials and Methods**

102 **2.1 WWTPs and sludge samples**

103 Sludge samples were obtained from the municipal WWTPs located in Librilla (W1),
104 Alhama de Murcia (W2) and Totana (W3) (Murcia, Spain) (Table S1).

105 **2.2 Microplastics (MPs) extraction procedure from sludge samples**

106 Three replicates of 20 mL sludge from each WWTP were introduced in 250 mL pre-
107 washed glass beakers. Afterwards, 40 mL of solution H₂O₂ (30 wt %) was added to the
108 beakers and left for 2 d to remove most of the organic matter present in the sample.
109 Then, MPs were extracted through density separation by adding 150 mL of a NaCl
110 saturated solution and stirring with a glass rod for 3 minutes. The solution was then
111 allowed to settle overnight. Afterwards, the supernatant was decanted and filtered. By
112 adding 150 mL of the saline solution, MPs decantation was facilitated by minimizing
113 resuspension of the settled particles.

114 This process was repeated four times. Since the density of the used saline solution
115 was 1.2 g cm⁻³, some polymers, mainly polyethylene terephthalate (PET; (density 1.29 –
116 1.40 g cm⁻³) and polyvinyl chloride (PVC; density 1.30 – 1.58 g cm⁻³) (Duxbury, 1992)
117 may not float in the solution but settle on the bottom of the beaker. To prevent this, in
118 the last repetition of the process, 6 g of sodium bicarbonate were added to the saline
119 solution. Then, the solution was heated for approximately 4 minutes until the
120 temperature reached between 45°C and 50°C and stirred with a glass rod to facilitate the
121 release of CO₂ bubbles that promote the flotation of MPs to the top of the saline
122 solution (per. obs.). The supernatant was filtered through glass fiber filters with a pore
123 size of 1.2 µm (Frisenette ApS, Knebel, Denmark).

124 Filters were left in petri dishes and the remaining organic matter was removed by
125 submerging them with approximately 20 mL of 1:1 (v/v) H₂O₂ (30 wt. %) + H₂SO₄ (96
126 wt. %) (Merck, Darmstadt, Germany) solution and left overnight. Since the H₂O₂:H₂SO₄
127 solution produces a very exothermic reaction, it was left to cool down before being used
128 to prevent possible damage to MPs. The used solution was chosen after several tests
129 using the following proportions of H₂O₂ (30 wt. %) and H₂SO₄ (96 wt. %): 1:1, 1:2 and
130 1:3 aiming to find the best compromise between organic matter removal and avoidance
131 of PET microfibers disintegration, which are the most sensitive component to be
132 disintegrated out of the main plastic debris found in the sludge (per. obs.). The best
133 results were obtained by using the 1:1 ratio in which fibers were not visually affected.
134 Higher proportions of H₂SO₄ lead to total or partial disintegration of the fibers.

135 After the exposition of filters to the acid solution, the liquid content in the petri
136 dishes was filtered. Both petri dishes and filters were rinsed off several times with
137 distilled water to ensure that no particles were retained. Then, particles in the filters
138 were identified by Raman spectroscopy and quantified. Since some particles were
139 retained on the walls of the bottom part of the funnel of the filtering equipment, after
140 each filtration, the funnel was separated from the main structure, washed with distilled
141 water and the liquid collected was filtered again in the same filtration unit to avoid MPs
142 losses.

143

144 **2.3 Recovery test**

145 A recovery test was performed to estimate the extraction efficiency and
146 reproducibility of the protocol used in this study. The recovery test was carried out
147 using commercially available plastic materials with different shapes and sizes (Table
148 S2) representative of the MPs found in sludge. The MPs were spiked into 20 mL of

149 sludge from W3. The particles used in the recovery test had specific colors and shapes
150 to facilitate their differentiation from particles already present in the sludge. In addition,
151 it was previously checked that the treatments applied to remove the organic matter did
152 not alter the particles used in the recovery test.

153 Fragments were composed of high density polyethylene (HDPE) and polypropylene
154 (PP) from a bleach bottle and a cap of a container, respectively. For films, HDPE and
155 low density polyethylene (LDPE) were used, from different plastic bags. The sizes of
156 fragments and films were: 0.4-0.6 mm, 0.8-1.0 mm, 1.9-2.1 mm and 2.4-2.6 mm. PET
157 microfibers were obtained from two T-shirts with a length of *ca.* 2 mm. Microbeads
158 were composed of butylene/ethylene copolymer (B/E) and they were obtained from a
159 shower gel presenting a size between 0.6 and 0.8 mm. Fragments, films and microfibers
160 were cut using a binocular magnifying glass (LFZ Dual, OPTECH, Ontario, Canada), a
161 graduated scale and a scalpel to produce the desired sizes. To test the repeatability, a
162 total of 10 particles of each size and type of material were added to 20 mL of sludge,
163 summing a total of 80 fragments, 80 films, 20 PET microfibers and 10 B/E microbeads.
164 The recovery test was performed six times to test its reproducibility, indicating the
165 calculated variation in the extraction efficiency when the measurements are reproduced.

166 The recovery efficiency percentage was calculated as follows:

$$167 \quad E (\%) = (P / Pr) \cdot 100$$

168 Where E stands for the Efficiency, P refers to the number of particles added and Pr
169 is the number of particles recovered.

170 **2.4 Characterization of MPs by Raman Spectroscopy**

171 MPs were characterized by Raman spectroscopy following the procedure indicated in
172 the Supplementary Data. Spectra of the suspected MPs were compared to the Raman

173 spectra of reference materials (Figure S1), (Boerio et al., 1976; Andreassen, 1999;
174 Long, 2004; Kida et al., 2016) (Table S3).

175 **2.5. Microplastics count**

176 The MPs present in each filter were counted using a binocular magnifying glass
177 (LFZ Dual, OPTECH, Ontario, Canada) with the aid of a transparent circular template
178 divided in eight sections that was placed at the top of the Petri dish.

179 **2.6. Quality control**

180 During the whole process of MPs extraction, blank filters were exposed to the air as
181 quality controls to evaluate possible cross-contamination (Woodall et al., 2015).
182 Particles found in these filters were analysed through Raman spectroscopy as explained
183 previously. See Supplementary Data for a more detailed explanation.

184

185 **2.7. Plant material and growth conditions**

186 The tomato plant (*Lycopersicon esculentum* Mill.) was chosen since it is widely
187 cultivated and shows low or no toxicity due to over fertilization with macronutrients
188 (Sainju et al., 2003). One-month tomato plants were transplanted outdoors on the 14th of
189 March of 2019 individually into a 3.3 L pot made of terracotta. The dimension of each
190 pot was 19 cm high, diameter of 20.7 cm at the top and 12.7 cm at the bottom. Five
191 soils, control (C), manure control (MC), and soils containing sludge of each WWTP
192 (W1, W2 and W3), were created using 50:50:0:0, 45:45:10:0, and 45:45:0:10
193 proportions of peat moss (COMPO SANA ® UNIVERSAL, Münster, Germany), silica
194 sand (grain size range: 0.4-0.8 mm), manure (FEMABÓN ®, Castellón, Spain) and
195 sludge, respectively.

196 Control soil was used as the reference substrate, while manure control soil was used
197 as a fertilizer-type amendment to soil aimed to simulate nutrient addition to the soil but
198 with an insignificant MPs concentration. To ensure that there was no nutrient deficiency
199 due to the different element composition of manure and sewage sludge in all soils, each
200 plant was fertilized monthly with 10 g of a solid slow-release fertilizer (NPK 12-8-18)
201 (Blueefficient Platinnum®, Salamanca, Spain), aiming to prevent changes in the
202 development of tomatoes due to a specific element deficiency while avoiding fertilizer
203 toxicity. Characteristics of the employed peat, manure, and fertilizer used are shown in
204 Table S4. Each type of soil was replicated in seven pots, each one containing a tomato
205 plant, so seven tomato plants were cultivated in each type of soil which made a total of
206 35 tomato plants.

207 The pots containing the plants were placed outdoors in an area of the campus of the
208 University of Alicante that had direct radiation from the sun during the whole day. A
209 wood stick was tied to each plant to prevent them from bending and a blue protective
210 mesh was placed in each pot to avoid plants from animal attacks. The experiment lasted
211 109 d. Each pot was watered with 500 mL of tap water. Watering frequency was once
212 every two days until June. Then, it was increased to twice per day until the experiment
213 finished on the 1st of July. The position of the pots was changed every month to avoid
214 possible effects due to placement.

215

216 **2.8 Measurement of pH and soil salinity**

217 Every month, pH and soil salinity were measured to monitor nutrient availability
218 and water absorption capacity of the tomato plants as it is described in Supplementary
219 Data.

220

221 **2.9. Collection of tomatoes**

222 The fruits of all tomato plants began to be harvested after three and a half months of
223 cultivation and the fruit selection was carried out according to their maturation status
224 with the purpose to collect tomatoes in a similar maturation stage, using as a criterion
225 the color of tomato fruits (Figure S2).

226 **2.10. Measurements of tomato plant growth parameters**

227 Once the experiment was concluded, tomato plants were taken out of the pots and
228 roots were cleaned in a cube with water to remove soil particles attached. Shoot length
229 of tomato plants was measured as the maximum length of the main stem, whereas root
230 length was determined as the maximum length of the longest root. Stem diameter was
231 measured with a digital vernier caliper (Mitutoyo CD-6 C5, Neuss, Germany). Then,
232 tomato plants were cut into the shoot and root parts and dried during 72 h at 70°C to
233 obtain the dry weight of each part.

234

235 **2.11. Statistical analysis**

236 The experiment consisted of a one factor (type of soil) design with five types of
237 soils: C, MC, W1, W2 and W3. Each treatment was replicated three times. Significant
238 differences among the response variables measured in the different types of soils were
239 analyzed through a one-way analysis of variance (ANOVA). Before ANOVA, the
240 normality was tested through Kolmogorov-Smirnov's test and heterogeneity of variance
241 of the data was tested using Cochran's test. If the data did not meet these assumptions,
242 transformations were applied, and assumptions were checked again. When significant
243 differences were found in the ANOVA main test, the *post-hoc* Tukey's test was used to
244 find which treatments significantly differed (Table S5). Statistical analyses were
245 performed with the R statistical software (v. 3.6.0; R Core Team, 2019). All statistical

246 tests were conducted with a significance level of $\alpha = 0.05$. Data were reported as mean
247 \pm standard error (SE).

248

249 **3. Results and discussion**

250

251 **3.1 Method extraction efficiency**

252

253 The mean average extraction efficiency for the total of particles spiked in the sludge
254 samples was $87 \pm 2\%$ (Figure 1). The range of particle size with the highest extraction
255 efficiency was 2.4-2.6 mm ($91 \pm 5\%$) whereas the lowest extraction efficiency was
256 obtained in particles with size in the range 1.9-2.1 mm ($87 \pm 3\%$). Fragments and films
257 were extracted with an average of $91 \pm 2\%$ and $86 \pm 3\%$, respectively. Microfibers had
258 a mean extraction efficiency value of $83 \pm 3\%$. Spiked microbeads showed $90 \pm 6\%$ of
259 extraction efficiency. As regarding the type of material PP, HDPE, LDPE and PET were
260 extracted with an efficiency of $91 \pm 2\%$, $89 \pm 2\%$, $88 \pm 3\%$ and $83 \pm 3\%$ respectively.

261 **Figure 1**

262 The obtained percentage values of extraction efficiency are, in general, comparable
263 with previous studies (Mahon et al., 2017; Sujathan et al., 2017; Hurley et al., 2018; Li
264 et al., 2018; Corradini et al., 2019). Only in some cases are slightly lower, which could
265 be due to the wide range of sizes of particles used in this study, generally reaching
266 lower sizes than other studies (Huerley et al., 2018). In addition, the recovery test
267 carried out in this study is comprehensively complete compared to previous ones where
268 only a few types of particles, materials and sizes were used (Mahon et al., 2017;
269 Sujathan et al., 2017; Hurley et al., 2018; Li et al., 2018;). Specifically, in this work the
270 recovery test uses microfibers, which is generally the main component of sludge despite
271 not being a particle type commonly used in recovery tests (Mahon et al., 2017; Sujathan
272 et al., 2017; Li et al., 2018). Our results indicate that this extraction method is capable

273 of efficiently recover PP, LDPE, HDPE and PET microfibers, which are the most
274 commonly used plastics (PlasticsEurope, 2019) and the most common types of MPs
275 found in sludge (Mahon et al., 2017; Li et al., 2018; Corradini et al., 2019). Thus, this
276 study demonstrates that the present extraction method is suitable for the extraction of
277 MPs in sewage sludge, while being inexpensive and safer than others that use other
278 salts, such as ZnCl and NaI.

279 **3.2 Characterization of microplastics in sludge samples**

280
281 The concentration of MPs was $17,870 \pm 2,174$; $27,821 \pm 1,357$ and $47,130 \pm 3,002$
282 particles kg^{-1} dry weight of sludge in W1, W2 and W3, respectively (Figure 2). The
283 sizes of the MPs were in the range of 0.31-2.11 mm (Figure S2).

284 **Figure 2**

285 W3 showed a significantly higher concentration of MPs in comparison with W1 and
286 W2. Concerning MPs shape, microfibers were the most abundant particle followed by
287 fragments, films and microbeads.

288 **Figure 3**

289 Microfibers represented the $93 \pm 2\%$, $89 \pm 5\%$ and $97 \pm 1\%$ of the total MPs in
290 W1, W2 and W3, respectively. All microfibers analyzed were made of PET, so it is
291 suspected that they were synthetic fibers used for acrylic garment, since microfibers in
292 sludge mainly come from cloth washing (Browne et al., 2011; Folkö, 2015; Åström,
293 2016). Additionally, microfibers, since they are made mostly of PET, their density,
294 higher than water and along with their high surface:volume ratio, they are prone to
295 precipitate in the sludge during wastewater processing in the WWTP. This explains that
296 PET microfibers were the most abundant MPs in sludge as previously reported (Mahon
297 et al., 2017; Li et al., 2018; Corradini et al., 2019).

298 Fragments accounted for the $6 \pm 2\%$, $9 \pm 4\%$ and $2 \pm 1\%$ of the total MPs in the
299 sludge from W1, W2 and W3, respectively. Films were uniquely found in W2 ($2 \pm 1\%$).
300 Fragments and films found were generally irregular. PE was the predominant polymer
301 in these types of particles, while PP showed a limited presence.

302 The lesser abundance of fragments and films in sludge in comparison to fibers sides
303 with previous studies (Mahon et al., 2017; Li et al., 2018; Corradini et al., 2019). This
304 may be due to PE, has lower density than water and it tends to float, and it is less likely
305 that their fate is the sludge. Additionally, because of its shape, fragments and films, for
306 a specific size, are more likely to be retained than microfibers in the pretreatment filters
307 of WWTPs before entering in the degreaser-desander channel. In fact, W2, which was
308 the WWTP with the largest mesh size pretreatment filter (70 mm) of all the WWTPs,
309 had the greatest percentage of fragments ($9 \pm 4\%$) and films ($2 \pm 1\%$) compared to the
310 percentage of fragments found in W1 ($6 \pm 2\%$) and W3 ($2 \pm 1\%$), which had a mesh
311 size of 2 and 3 mm, respectively. These findings seem a plausible explanation of the
312 low presence of fragments and films in sludge. Microbeads only constituted 0.4 ± 0.4
313 %, $0.4 \pm 0.4\%$ and $0.2 \pm 0.2\%$ of the total MPs in W1, W2 and W3, respectively as
314 previously reported (Mahon et al., 2017; Li et al., 2018).

315 As regards the extraction process in the sludge samples, out of the total particles
316 extracted, $61 \pm 4\%$, $19 \pm 4\%$, $13 \pm 2\%$ and $7 \pm 2\%$ of the particles were recovered in
317 the first, second, third and fourth extraction step, respectively. In the last extraction step,
318 bubbles produced by the bicarbonate kept microfibers on the surface of the solution
319 used for the separation of MPs through difference of density. Thus, the use of
320 bicarbonate for MPs extraction could be advisable to maximize the process, especially
321 in matrices with high microfibers content.

322 The average concentration of MPs in the sludge amongst the three WWTPs was
323 $30,940 \pm 8,589$ particles kg^{-1} dry weight, being higher than previous studies (Corradini et
324 al., 2019); (van den Berg et al., 2020); (Mahon et al., 2017).

325 3.3. Measurements of tomato plant parameters

326
327 After 109 d growth, significant differences among soil treatments were found in
328 biomass production, but not in the morphological variables studied (Figure 4).

329 Figure 4

330 Shoot biomass was significantly higher in W2 (79.2 ± 7.3 g) than in C (35.8 ± 4 g),
331 MC soil (41.4 ± 3.4 g) and W3 (53.4 ± 5.1 g). Significant differences were also found
332 between W1 (60.9 ± 7.2 g) and C samples. For root biomass, W2 (67.8 ± 7.3 g) was
333 significantly higher than C (23.4 ± 5.1 g), MC (36 ± 10.4 g) and W3 (32.4 ± 5.1 g).
334 Total biomass was significantly higher in W2 (146.9 ± 11.3 g) than in C (59.1 ± 8.7 g),
335 MC (77.4 ± 11.8 g) and W3 (85.8 ± 11.2 g) soils. Significant differences were found
336 between W1 (60.9 ± 7.2 g) and C samples. The values of stem diameter, height and root
337 length ranged between 11.7 ± 1.5 and 10.3 ± 1.1 mm, 74.0 ± 5.4 and 61.9 ± 15.4 cm and
338 34.2 ± 6.0 and 27.4 ± 6.8 cm, respectively.

339 The number of tomatoes harvested on each tomato plant did not significantly vary as
340 regards the type of soil. MC soil had the highest tomatoes production with 5 ± 1
341 tomatoes while W3 produced less than 2 fruits. During the experiment, the number of
342 mature tomatoes produced in plants grown in C and MC soil was significantly higher
343 (1.4 ± 0.3 and 1 ± 0.4 tomatoes, respectively) than the ones grown in soils containing
344 sludge with MPs (W1; $0,13 \pm 0,13$; W2: $0,14 \pm 0.14$; W3: 0) (Figure 5).

345 Figure 5

346 At the end of the experiment, MC had the highest percentage of plants (75%) that
347 produced tomatoes, whereas in C or soils containing sludge from W1, W2 and W3 this
348 percentage was lower being 50, 37.5, 25 and 25%, respectively.

349 The mean values of pH obtained for W1 (6.4 ± 0.1) and W3 (6.5 ± 0.1) were
350 considered slightly acidic, whereas W2 (6.6 ± 0.4), MC (7.2 ± 0.1) and C (6.8 ± 0.1) were
351 considered neutral (Juárez et al., 2004), which limits nutrient deficiency problems of
352 nutrient availability related to highly acidic or basic soils (Sainju et al. 2003). The mean
353 values of electrical conductivity obtained from C (3.3 ± 0.2 dS/m), W1 (2.9 ± 0.2
354 dS/m), W2 (2.8 ± 0.1 dS/m) and W3 (2.7 ± 0.1 dS/m) were considered very slightly
355 saline (2-4 dS/m) whereas MC soil was considered slightly saline (5.3 ± 0.2 dS/m)
356 (Schoeneberger et al., 2002). The sludge addition to soils generally results in an
357 increase of their salinity, but in case of saline soils, it can reduce their salinity (Pomares
358 et al. 1998). Peat moss, that was used as the base of the soil component in the present
359 experiment, can retain a large amount of cations, which could be responsible of
360 increasing the levels of salinity. The variety of tomato plants used, Raf, have a threshold
361 of tolerance to salinity of 6 dS/m (Nelson, 2011). Thus, the levels of salinity in the
362 different types of soil is not expected to negatively affect the growth and yield of tomato
363 plants.

364 Nutrient levels in the different sludge used were in some cases notably different. For
365 example, N was $7.6 \pm 0.9\%$ in W2, while in W1 and W3 was 1.9 ± 0.2 and 5.4 ± 0.6 ,
366 respectively (Table S1). High contents of N concentration can promote biomass at the
367 expense of fruit yield (Sainju et al. 2003). Our results indicate a notable shoot and root
368 growth of tomato plants in soil W2, which was significantly different to the plants
369 grown in soil W3, but not in W1, which had the lowest N content (Fig. 4). Nevertheless,

370 fruit production was not affected in plants grown in any of the soils treated with sludge
371 (Fig. 5).

372 Phosphorous and K values were also different among the used sludges. Therefore, in
373 our experiment a solid slow-release fertilizer containing N, P and K was applied to all
374 the treatments aimed to prevent nutrient limitation. Other macronutrients, such as Mg
375 and Ca, despite the content being different among sludges, the levels were high enough
376 that no deficiency was expected. Nutrient toxicity due to the above-mentioned nutrients
377 is not expected in tomato plants, apart from the commented issue with N (Sainju et al.
378 2003).

379 Our results suggest contrasting effects of sludge addition in soils since the biomass
380 production was higher in plants cultured with sludge than the ones cultured in the C or
381 MC soils. However, tomato plants cultured in soils treated with sludge from W3, which
382 had the highest concentration of MPs, had the lowest biomass and tomato production,
383 while it did not produce any mature tomato during the experiment. Plant biomass can be
384 influenced by many factors, such as soil humidity, soil and air temperature,
385 photoperiod, solar radiation, precipitations, genotype, etc. One of the most important
386 factors influencing biomass is soil nutrient availability since both nutrient deficiency
387 and toxicity negatively affect total biomass and fruit production (Msilini et al., 2009;
388 Karim et al., 2012). The solid slow-release fertilizer aimed to prevent nutrient
389 limitation. The shoot:root biomass ratio of cultured plants did not show significant
390 differences among plants cultured in the different soils, suggesting non-relevant
391 differences in nutrient availability among soils (Ericsson, 1995). Water was
392 homogeneously provided, while it was ensured that all the plants were similarly
393 exposed to light.

394 The effects of MPs in soil have been scarcely studied, but are expected to be highly
395 influenced by the shape (Rilling et al., 2019) and size (Bosker et al., 2019), being
396 microfibers among the type of MPs that is expected to have the strongest effects (de
397 Souza Machado et al., 2019). In the present experiment, differences in plant biomass
398 among soils with sludge were noticeable. Root mass is expected to increase in the
399 presence of microfibers since they can lower soil bulk density (de Souza Machado et al.,
400 2018) promoting increased root growth due to reduced penetration resistance for plant
401 roots and improved soil aeration (de Souza Machado et al., 2019). These differences
402 could also directly and indirectly affect soil structure through the modification of soil
403 aggregation (de Souza Machado et al., 2018). Our results side with previous findings
404 showing the largest biomass in the root, as well as, in shoot in soils with sludge,
405 especially in W2 (de Souza Machado et al., 2018; de Souza Machado et al., 2019). Very
406 high concentrations of MPs, as in W3, can have the opposite effect, reducing root
407 biomass production (Jiang et al., 2019).

408 Our results indicate a lower crop in the tomato plants grown under soil treated with
409 sludge at the time of the end of the experiment. This could be because of microfibers in
410 increasing C:N ratio as a consequence of the modification of nutrient availability
411 derived from alterations in water dynamics reported in onion plants (de Souza Machado
412 et al., 2019). Nutrient alterations can lead to plant stress, lowering crop production (Li
413 et al., 2009). Since the experiment did not last until the end of the crop season, we could
414 not know if the tomato plants suffered a reduction in crop or only a delay in crop
415 production. Nevertheless, a delay in the crop production is expected to lower the overall
416 crop production that is possible to be harvested (Thomison et al., 2011).

417 When interpreting the results of this study, possible variables that were not
418 controlled, such as nutrient level, could influence them and need to be taken into

419 account. Despite the solid slow-release fertilizer provided to all the treatments aimed to
420 prevent nutrient limitation, nutrient concentration was not equal in the different
421 treatments. Further experiments, aiming to have a more similar nutrient concentration
422 among treatments would be desirable.

423 Additionally, sewage sludge is a complex matrix that does not only contain MPs,
424 but also metals, pathogens and organic toxicants. The levels of metal concentration
425 found in sludge samples were below the ones established in the European Council
426 Directive 86/278/EEC (Table S1). Sludge samples also contained bacteria such as
427 *Escherichia coli* and *Salmonella sp.*, that inhabit human intestinal tracts (Table S1) and
428 so are not necessarily pathogens, but certain strains can cause pathogenicity. Thus,
429 nutrients, metals, bacteria and other variables, as well as the interactions of these
430 variables with MPs could have influenced the results of this study. Because the above
431 commented variables are not controlled, the experimental design of our study using
432 different sludges does not allow us to demonstrate a direct cause-effect of MPs.
433 However, this work allows us to test the real effects of sewage sludge containing MPs.
434 Complementary studies would be needed to increase the insight on the possible effects
435 of MPs from sewage on agriculture.

436 This study suggests, that despite the large number of MPs reported in sludge (Lusher
437 et al., 2017; Mahon et al., 2017; Li et al., 2018; Corradini et al., 2019; van den Berg et
438 al. 2020), these estimates could underestimate real concentrations or indicate that the
439 number of MPs is growing. Under this scenario of high amounts of MPs exported
440 through sewage sludge to the environment (De Souza Machado et al. 2018), it is urgent
441 that policy makers define maximum levels of MPs in sludge used for agriculture as it
442 has already been done with other toxic compounds, such as metals (86/278/EEC).

443 To the best of our knowledge, no study has tested the effects of sludge with MPs in
444 the yield of agriculture crops. Our results show that sewage sludge with large
445 concentrations of MPs can notably affect tomato plants, which could be due to a greater
446 or lower extent to MPs. However, other possible factors may have been responsible or
447 can produce additive or interactive effects with MPs. Thus, further work is necessary to
448 corroborate these findings and to explain the mechanisms of possible effects of MPs on
449 plants, such as tomato.

450 **Author Contributions:**

451 Conceptualization, C.S., A.B.; methodology R.H., C.S., A.B., P.N.; formal
452 analysis, R.H., A.B., C.S.; ; investigation, R.H., P.N., A.B., C.S.; ; resources, A.B., C.S.;
453 data curation, A.B., C.S., R.H.; writing—original draft preparation, R.H., A.B., C.S.;
454 writing—review and editing, C.S., A.B.; ; supervision, A.B., C.S. All authors have
455 read and agreed to the published version of the manuscript.

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458 **Declaration of competing interest**

459 The authors declare that there are no conflicts of interest

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466

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613 FIGURES

614 Fig. 1. Percentage of particles extracted in the recovery test (mean \pm SE; n=2)
615 concerning size, type of polymer and particle.

616 Fig. 2. Number of microplastics (mean \pm SE; n=3) found in sludge samples from the
617 wastewater treatment plants in the cities of Librilla (W1), Alhama de Murcia (W2) and
618 Totana (W3) in Spain. Different letters on top of the bars represent statistically
619 significant differences (Tukey-HSD test, $p < 0.05$).

620 Fig. 3. Percentage of types of polymers (A) and particles (B) (mean \pm SE; n=3) found in
621 the cities of Librilla (W1), Alhama de Murcia (W2) and Totana (W3) in Spain.

622 Fig. 4. Morphological and biomass parameters (mean \pm SE; n=7) after 109 d of growth..
623 Different letters on top of the bars represent statistically significant differences (Tukey-
624 HSD test, $p < 0.05$). Control (C), Manure Control (MC), Librilla (W1), Alhama de
625 Murcia (W2), Totana (W3).

626 Fig. 5. Number of tomatoes (A) and number of mature tomatoes produced (B) (mean \pm
627 SE).. Different letters on top of the bars represent statistically significant differences
628 (Tukey-HSD test, $p < 0.05$). Control (C), Manure Control (MC), Librilla (W1), Alhama
629 de Murcia (W2), Totana (W3).

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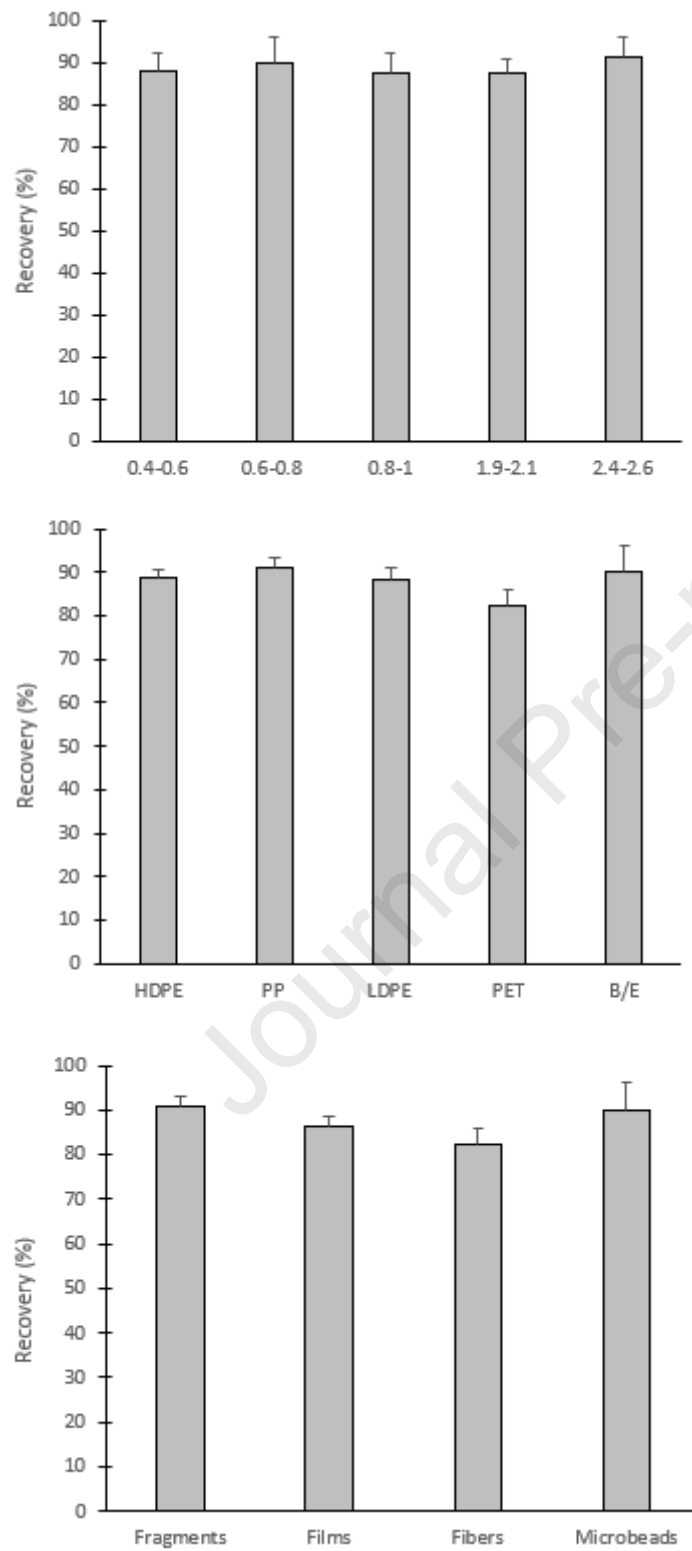
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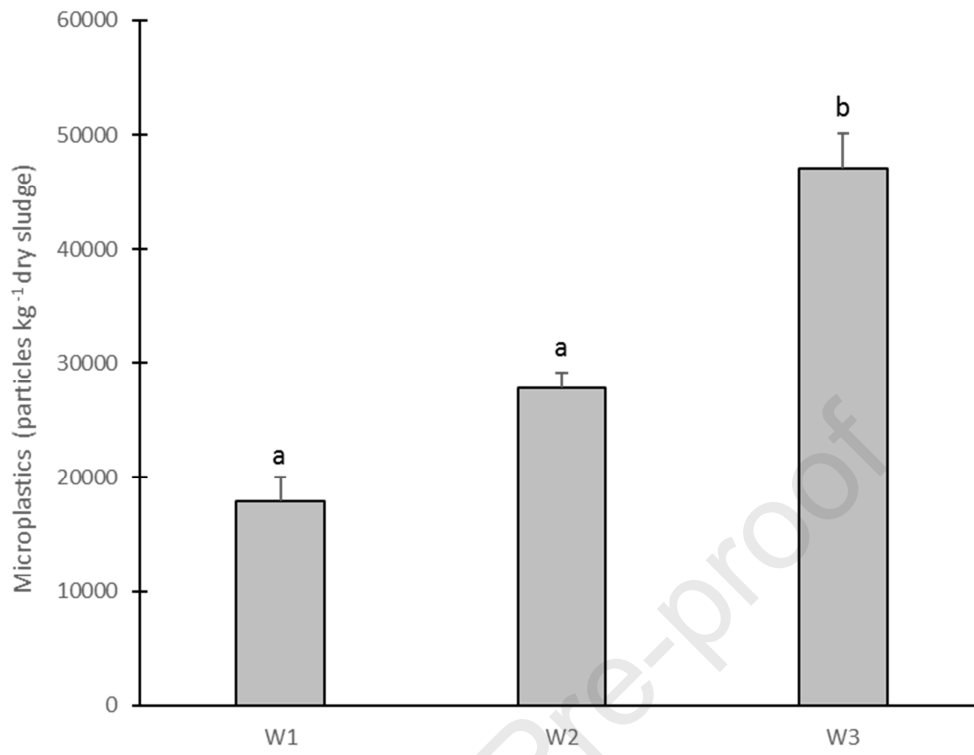
638 Figure 1



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641 Figure 2

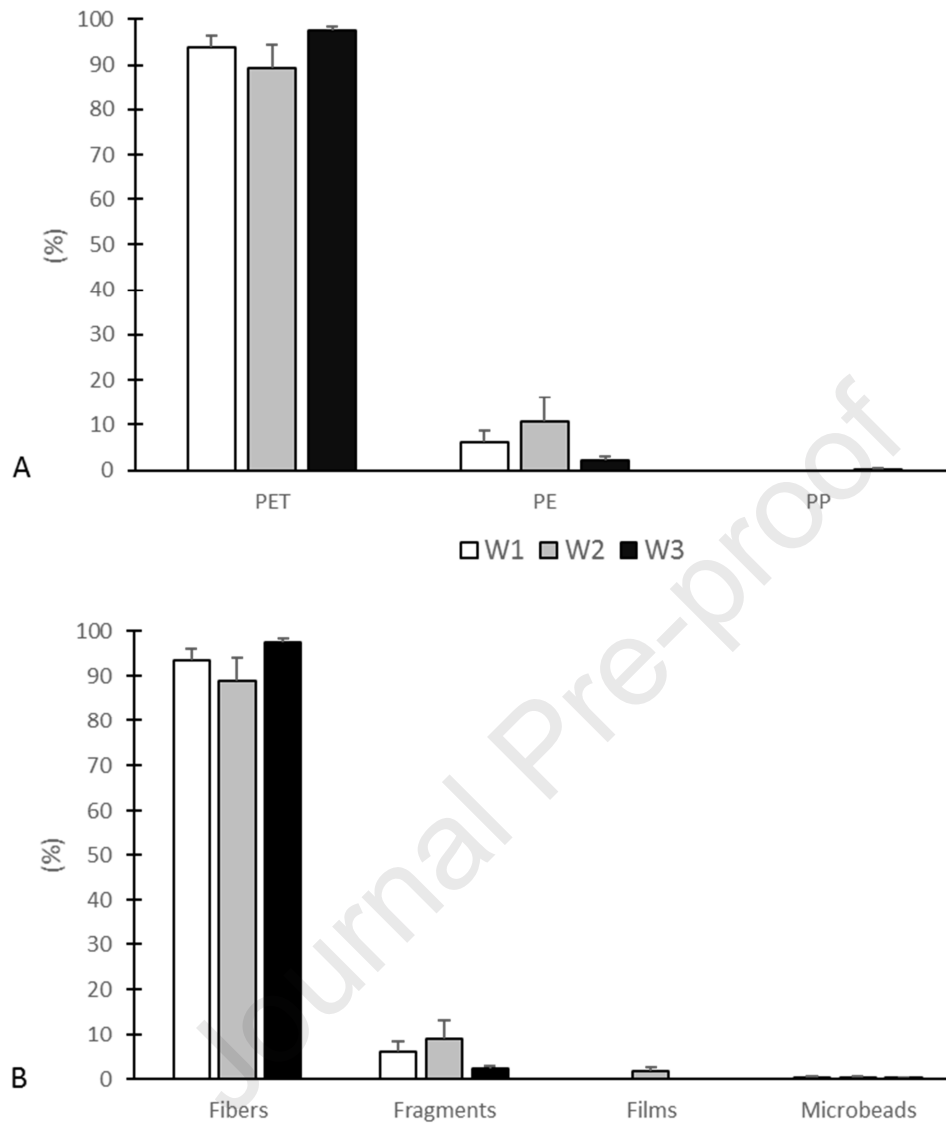


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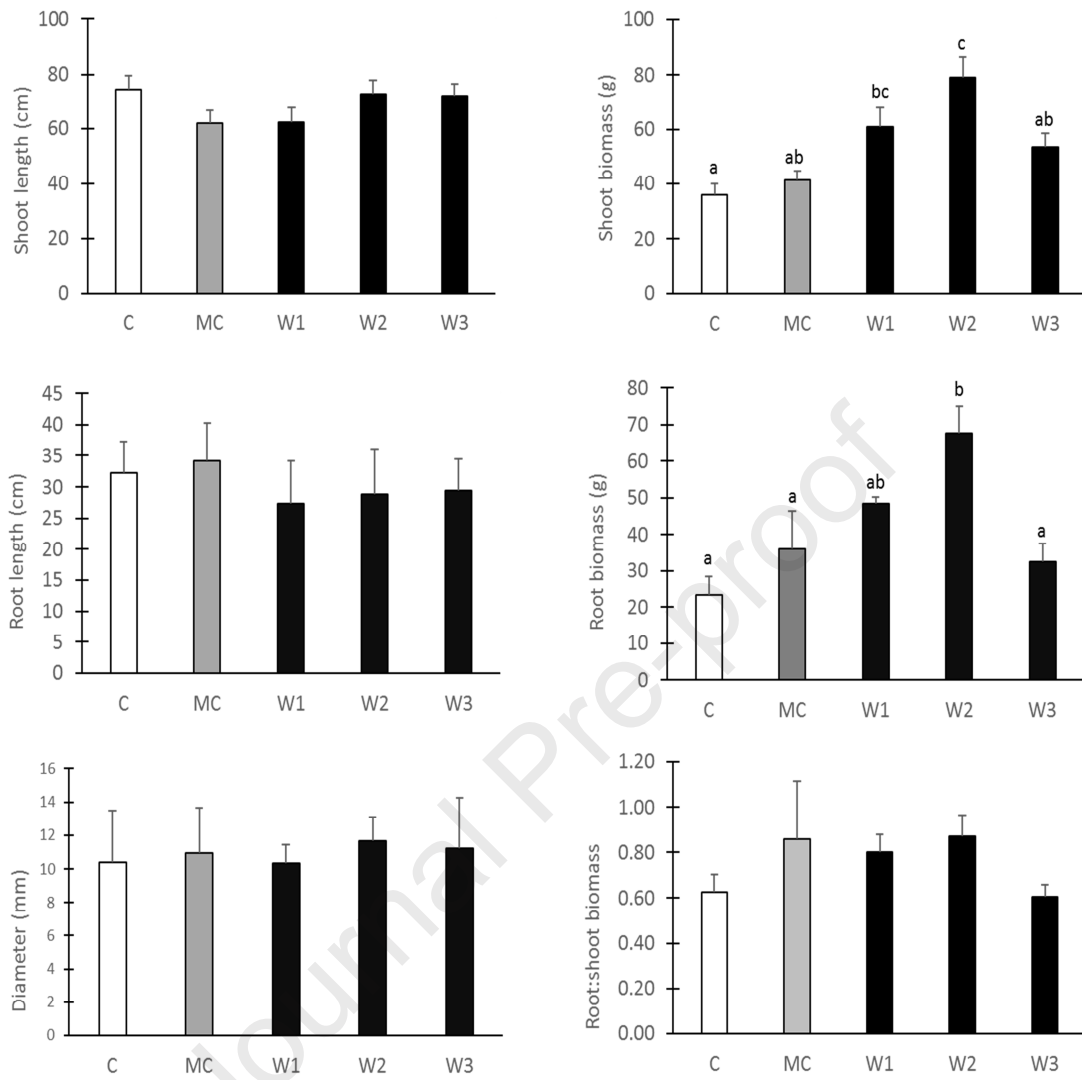
Figure 3



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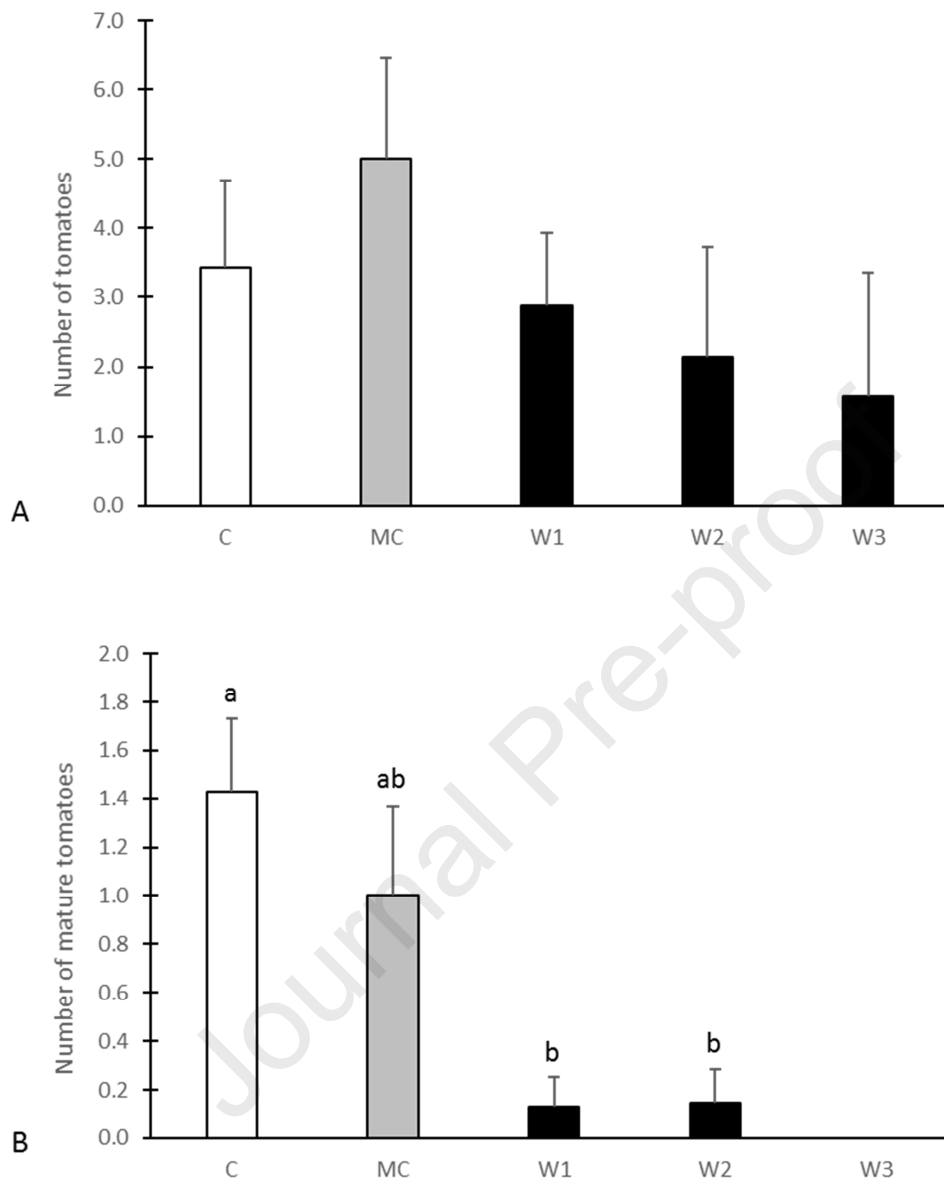
647 **Figure 4**



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651 **Figure 5**

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654 **SUPPLEMENTARY DATA**655 **2.4. Characterization of MPs by Raman Spectroscopy**

656 MPs were characterized by using a Raman spectrometer NRS-5100 (Jasco, Madrid,
657 Spain) equipped with LMU-20X-UVB lens. The laser excitation frequency and intensity
658 used was 784.79 nm and 11.8 mW, respectively. Raman spectra were recorded with a
659 charge-coupled device camera (UV-NIR range, 1024 × 255 pixels) electrically cooled to
660 -70°C. Raman spectra were obtained between 162 and 1886 cm⁻¹ with a spectral
661 resolution of 2.47 cm⁻¹.

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663 **2.6. Quality Control**

664 A total of 29 fibers were found in 3 blank filters. The mean concentration of MPs
665 (9.7 ± 0.3 particles) in blank filters were subtracted to the mean value of the extracted
666 MPs.

667 To test for modifications in the Raman spectra of the samples due to the chemical
668 agents employed to remove the organic matter, a preliminary trial was performed with
669 all the types of polymers used in the recovery test. The Raman spectra of all the types of
670 polymers were obtained before and after performing the extraction protocol performed
671 to the sewage sludge samples. No relevant changes in the intensity of the characteristic
672 bands (Figure S3) were found in comparison with the spectra of the same materials
673 before applying the extraction protocol.

674 To minimize contamination during the lab procedure all the containers were covered
675 with aluminum foil, lab instrumentals were washed before use and were made of glass,
676 the windows of the laboratory were closed, and 100% cotton coats were worn.

677 The Raman analysis of the fiberglass, the material the filter was made of, did not
678 emit fluorescence that could interfere with the analysis of the samples, before and after
679 applying the treatment to remove organic matter.

680 **2.8. Measurement of pH and soil salinity**

681 To determine pH values, air-dried soil samples from the pots were sieved through a
682 2 mm mesh to keep all primary soil particles and remove roots and bigger soil
683 aggregates. Then, 40 mL of sieved soil was added to a glass beaker and slowly wetted
684 with distilled water (*sensu* Juárez et al., 2004). After that, soil was stirred for 20
685 seconds with a glass rod and left 60 minutes before the pH was measured (pH Basic 20,
686 Crison Instruments S.A., Barcelona, Spain). To monitor soil salinity, a container was
687 placed under each tomato plant to collect 50 mL of the water lixiviated after watering
688 the plants. Then, the electrical conductivity of the collected water samples was
689 measured by using an electrical conductivity meter (Crison Basic 30, Barcelona, Spain).

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704 Table S1. WWTPs and sludge samples characteristics.

705 Table S2. Characteristics of MPs used in the recovery test. Number of particles shown
706 are referenced to one recovery test.

707 Table S3. PE, PET and PP Raman bands utilized for the identification of the
708 microplastics.

709 Table S4. Label information of the products utilized in the composition of the soil
710 treatments.

711 Table S5. Results of the ANOVA for the SL (shoot length), RL (root length), D
712 (diameter), SB (shoot biomass), RB (root biomass), RB:SB (root:shoot biomass ratio),
713 NT (number of tomatoes), NMT (number of mature tomatoes) and MPA (microplastics
714 abundance). The factors were *WWTPs* (wastewater treatment plants) and *TOS* (type of
715 soil).

716 Figure S1. Raman spectrum of a fiber and its similarity to a reference material.

717 Figure S2. Photographs of some microplastics found in the sewage sludge samples.

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719 Figure S3. Tomato appearance and color at harvest time.

720 Figure S4. Raman spectra of a PP fragment (A), LDPE film (B) and PET fiber (C)
721 before (grey line) and after (black line) the reagents used for removing the organic
722 matter.

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729 **Table S1.** Characteristics of the wastewater treatment plants (WWTP) and sludge. W1, W2 and W3,

730 correspond to the three WWTPs located in Librilla, Alhama de Murcia and Totana, respectively in

731 Murcia, Spain.

	W1	W2	W3
WWTP			
Population equivalent	4,312	17,293	27,628
Type of treatment	Secondary and tertiary	Secondary and tertiary	Secondary and tertiary
Water sources	Domestic	Domestic and industrial	Domestic and industrial
Treatment capacity (m ³ /year)	227,815	994,596	1,684,947
Sludge production (kg dry sludge /year)	55,632	317,280	695,592
Sludge			
Dry matter (%)	18±2	13±1	18±2
Total organic matter (%)	77±6	84±7	66±5
N (%)	1.9±0.2	7.6±0.9	5.4±0.6
P (P ₂ O ₅) (mg kg ⁻¹)	54,204±10,841	36,846±7,369	37,716±7,543
K (K ₂ O) (mg kg ⁻¹)	9,220±1,660	3,484±627	6,570±1,183
Mg (MgO) (mg kg ⁻¹)	15,036±2,707	10,384±1,863	15,692±2,825

Ca (CaO) (mg kg ⁻¹)	31,906±6,381	31,668±6,334	70,714±14,143
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734 **Continuation Table S1**

	W1	W2	W3
Sludge			
Cu (mg kg ⁻¹)	189±28	186±28	449±67
Cd (mg kg ⁻¹)	<2.0	<2.0	<2.0
Cr (mg kg ⁻¹)	27±5	21±4	32±6
Hg (mg kg ⁻¹)	0.23±0.08	0.24±0.08	0.27±0.09
Ni (mg kg ⁻¹)	20±3	16±2	19±3
Pb (mg kg ⁻¹)	18±3	25±5	40±7
Fe (mg kg ⁻¹)	4689±750	3850±616	8223±1316
Zn (mg kg ⁻¹)	511±87	608±103	1599±272
<i>Escherichia coli</i> (CFU/g; confidence interval)	140000±88000- 220000	140000±88000- 220000	17000±11000- 27000
<i>Salmonella sp.</i> (presence/25g)	absence	presence	presence

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744 **Table S2.** Characteristics of MPs used in the recovery test. Number of particles shown
 745 are referenced to one recovery test.

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Polymer type	Number of particles	Particle shape	Color	Source	Size (mm)
HDPE	40	Fragment	Blue	Bleach bottle	0.4-0.6, 0.8-1, 1.9-2.1, 2.4-2.6
HDPE	40	Film	Black	Plastic bag	0.4-0.6, 0.8-1, 1.9-2.1, 2.4-2.6
LDPE	40	Film	Blue	Plastic package	0.4-0.6, 0.8-1, 1.9-2.1, 2.4-2.6
PP	40	Fragment	Red	Bottle tap	0.4-0.6, 0.8-1, 1.9-2.1, 2.4-2.6
PET	10	Fiber	Blue	100% polyester T-shirt	2
PET	10	Fiber	Yellow	100% polyester T-shirt	2

B/E	10	Microbead	Blue	Palmolive Thermal Spa Mineral Massage ®	0.6 – 0.8
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Table S3. Examples of the Raman bands utilized for the identification of PE, PET and PP.

Polymer	Wavenumber (cm ⁻¹)	Assignment	Reference
PE	1063	Anti-symmetric stretching (C-C)	Kida et al., (2016)
	1080	Stretching (C-C)	
	1130	Symmetric stretching (C-C)	
	1298	Twisting (C-C)	
	1440	Bending (CH ₂)	
	1460	Bending (CH ₂)	
PET	1290	Stretching C(O)-C	Boerio et al., (1976)
	1414	CCH bending and OCH bending	
	1615	Ring mode 8a	
	1726	Stretching C=O	

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Continuation Table S3

Polymer	Wavenumber (cm ⁻¹)	Assignment	Reference
PP	809	Rocking CH ₂ , Stretching CC _b , Stretching C-CH ₃	Andreassen (1999)
	841	Rocking CH ₂ , Stretching CC _b , Stretching C-CH ₃ , rocking CH ₃	
	900	Rocking CH ₃ , rocking CH ₂ , bending CH	
	941	Rocking CH ₃ , stretching CC _b ,	
	973	Rocking CH ₃ , stretching CC _b ,	
	998	Rocking CH ₃ , bending CH, wagging CH ₂	
	1040	Stretching C-CH ₃ , Stretching CC _b , bending CH	
	1152	Stretching CC _b , stretching C- CH ₃ , bending CH, Rocking CH ₃	
	1219	Twisting CH ₂ , bending CH, stretching CC _b	

1330	Bending CH, twisting CH ₂
1360	Symmetric bending CH ₃ , bending CH
1458	Asymmetric bending CH ₃ , bending CH ₂

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766 **Table S4.** Label information of the products utilized for the composition of the soil

767 treatments.

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Product	Label information
COMPO SANA ® UNIVERSAL	Composition: black peat, Perlita, Agrosil ® and lime; pH (CaCl ₂): 5,0 – 6,5; Salt content (KCl) g/l: <3; Content in subscription (soluble nutrients): 200 – 400 mg/l Nitrogen (N), 200 – 500 mg/l Phosphate (P ₂ O ₅), 300 – 500 mg/l Potassium oxide (K ₂ O)
Manure FEMABÓN ®	Composition: herbaceous peat and compost manure; organic material (dry weight): 48%; bulk density: 610 g/l; electrical conductivity: 3 dS/m pH: 7.9
BLUEFFICIENT PLATINNUM®	12% Nitrogen; 7,5% (N) ammoniacal; 4.5% (N) ureic; 8% P ₂ O ₅ ; 18% K ₂ O; 2% MgO 17% SO ₃ ; 0.1% Mn; 0.1% Zn 3%; pH 6.0 inhibitor

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800**Table S5.** Results of the ANOVA for the SL (shoot length), RL (root length), D (diameter), SB (shoot biomass), RB (root biomass), RB:SB (root:shoot biomass ratio), NT (number of tomatoes), NMT (number of mature tomatoes) and MPA (microplastics abundance). The factors were WWTPs (wastewater treatment plants) and TOS (type of soil).

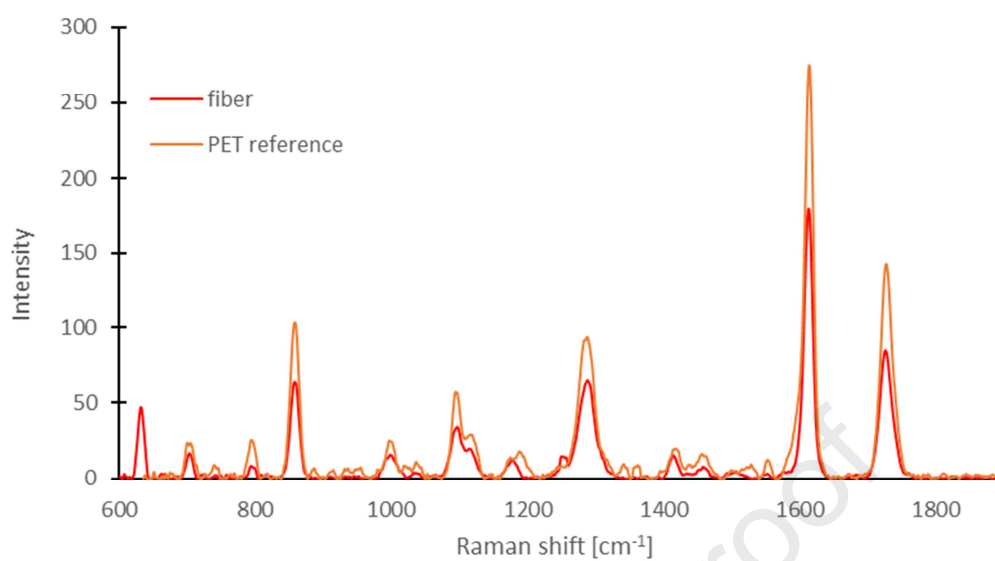
Source of variation	d.f.	SL			RL			D			SB		
		MS	F	P	MS	F	P	MS	F	P	MS	F	P
TOS	4	245.4	1.574	>0.2	51.85	1.507	>0.2	2.261	0.843	>0.5	2020	8.351	<0.001
Residual	30	155.9			34.40			2.681			242		
Total	34												
Cochran's C test			C = 0.30043, P > 0.05			C = 0.39329, P > 0.05			C = 0.49131, P < 0.05			C = 0.36118, P > 0.05	
Transformation			None			None			None			None	
Tukey's test												W2 > C	>0.0001
												W2 > MC	>0.001
												W2 > W3	>0.03
												W1 > C	>0.03

Source of variation	d.f.	RB			RB:SB			NT			NMT		
		MS	F	P	MS	F	P	MS	F	P	MS	F	P
TOS	4	2065.5	6.041	<0.01	0.1124	1.105	>0.3	8.896	0.513	>0.7	0.7198	4.755	<0.01
Residual	30	341.9			0.1017			17.343			0.1514		
Total	34												
Cochran's C test			C = 0.37135, P > 0.05			C = 0.69274, P < 0.05			C = 0.30208, P > 0.05			C = 0.43627, P > 0.05	
Transformation			None			None			None			logarithmic	
Tukey's test			W2 > C	>0.0008								C > W1	>0.01
			W2 > MC	>0.03								C > W2	>0.02
			W2 > W3	>0.009								C > W3	>0.005

Source of variation	d.f.	MPA		
		MS	F	P
WWTPs	2	663975790	34.97	<0.001
Residual	6	18987671		
Total	8			
Cochran's C test			C = 0.47455, P > 0.05	
Transformation			None	
Tukey's test			W3 > W1	> 0.03
			W3 > W2	> 0.0004

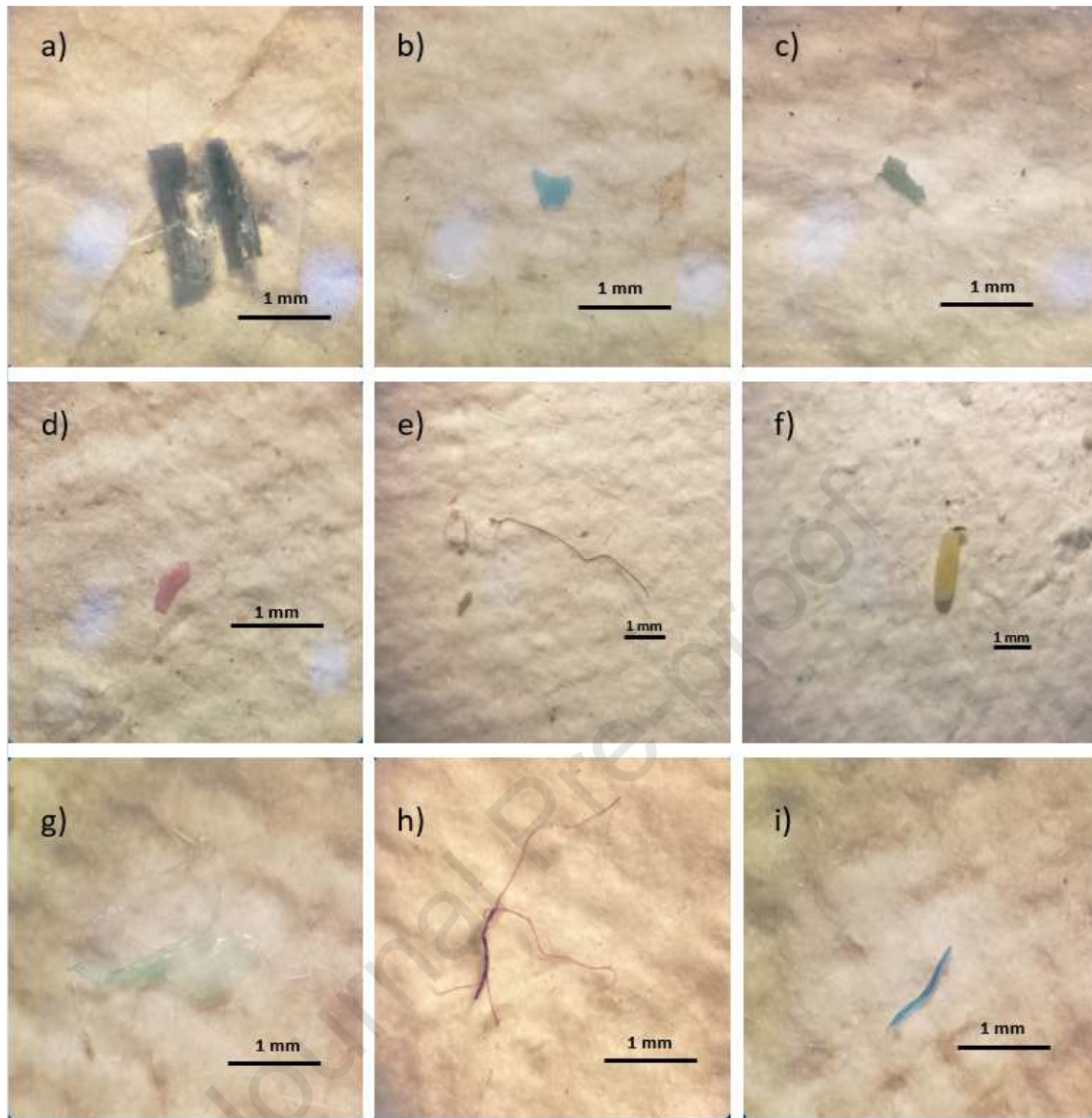
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Fig. S1. Raman spectrum of a fiber and its similarity to a reference material.



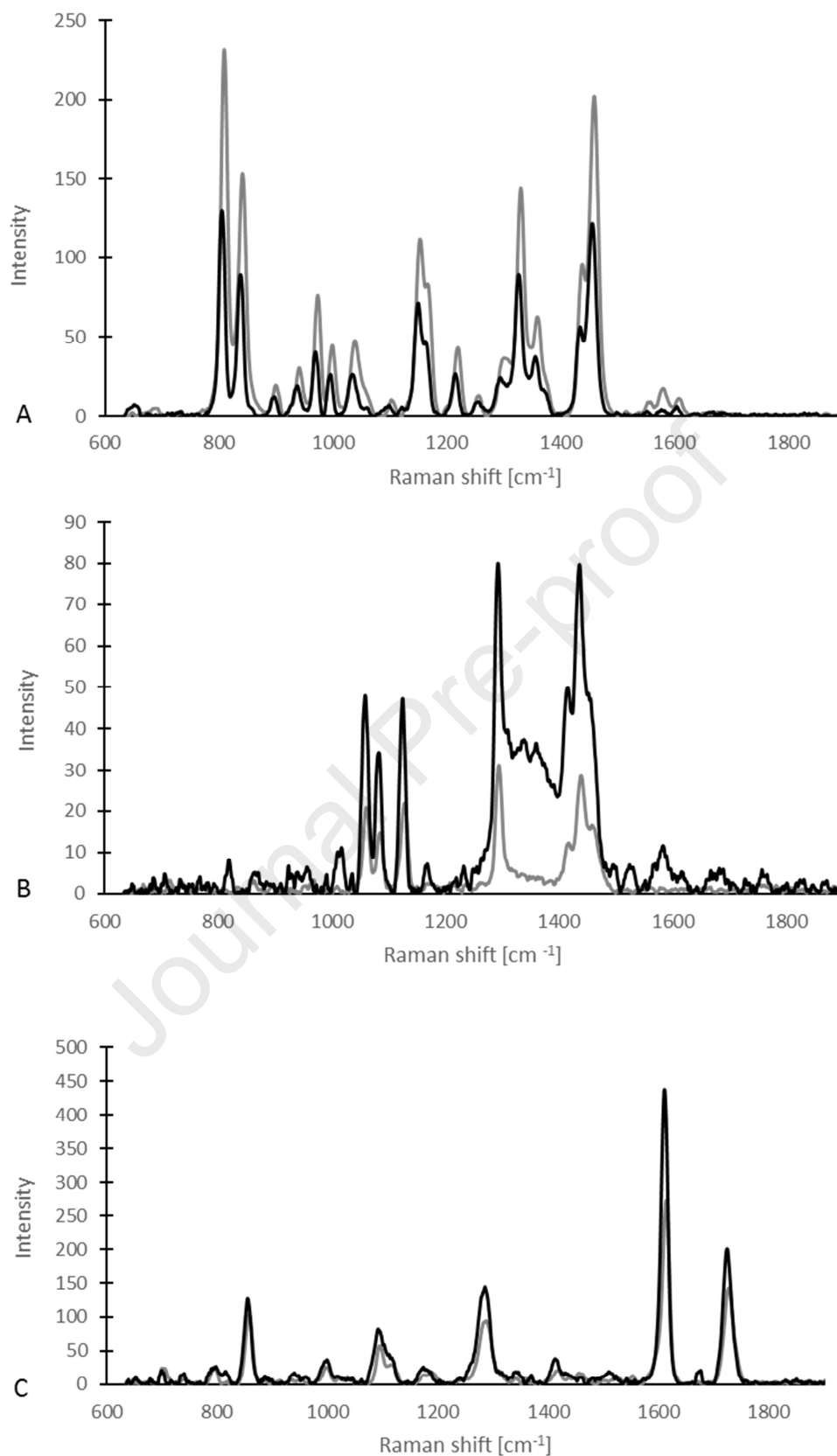
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844 **Fig. S2.** Photographs of some microplastics found in the sewage sludge samples.
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Fig. S3. Tomato appearance and color at harvest time.



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Fig. S4. Raman spectra of a PP fragment (A), LDPE film (B) and PET fiber (C) before (grey line) and after (black line) the exposition to the reagents used for removing the organic matter.

HIGHLIGHTS

An extraction protocol for MPs from sewage sludge has been validated.

Up to $31,000 \pm 8,600$ particles kg^{-1} dry weight in sewage sludge were estimated.

Sewage sludge containing MPs fostered the growth of tomato plants.

Sewage sludge containing MPs delayed and diminished fruit production.

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Author Contributions:

Conceptualization, C.S., A.B.; methodology R.H., C.S., A.B., P.N.; formal analysis, R.H., A.B., C.S.; ; investigation, R.H., P.N., A.B., C.S.; ; resources, A.B., C.S.; data curation, A.B., C.S., R.H.; writing—original draft preparation, R.H., A.B., C.S.; writing—review and editing, C.S., A.B.; ; supervision, A.B., C.S. All authors have read and agreed to the published version of the manuscript.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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